

Reference: DSHB3051

A.B.E. - Technical Data Sheet

**Product: Listeria Oxford Selective Supplement** 

# **Specification**

Sterile selective supplement used for Listeria isolation in food samples.

#### Presentation

**Shelf Life** Storage **Packaging Details** 10 Freeze dried vials Vial 49 months 23x60 mm glass vials, tag labelled, White plastic cap -2-25 °C 10 vials per box. with: 6 ± 0.1 g

#### Composition

Compositon (g/vial) Note: Each vial is enough to supplement 500 ml of Oxford medium Base.

Cyclohexymide	0.2000
Colistin sulphate	
Acriflavine	
Cefotetan	0.0010
Phosphomycin sodium salt	0.0050

Reconstitute the original freeze-dried vial by adding:

Sterile solvent (50% Ethanol/water)......9 ml

## **Description / Technique**

#### Description:

Listeria Selective Supplement (Oxford Formulation) is added to Oxford agar base in order to obtain a complete selective medium for the detection of Listeria monocytogenes from clinical and food specimens.

Listeria monocytogenes plays an important role in human and animal illness and the sources of infections are numerous.

In the last years the lack of an effective selective medium has been a gap in the detection of Listeria, as it can be easily and completely overgrown by competing flora.

With his supplement, which supplies the selective inhibitory components acriflavine, colistin sulphate, cefotetan, cycloheximide and fosfomycin, the competing flora is inhibited. Listeria monocytogenes is differentiated becasue it hydrolyses aesculin, producing black zones around the colonies .

Gram-negative bacteria are completely inhibited and also most of the unwanted Gram-positive species. Some strains of enterococci grow poorly and exhibit a weak aesculin reaction, usually after 40 hours incubation. Some staphylococci may grow as aesculin-negative colonies.

#### Technique:

Collect, dilute and prepare samples and volumes as required according to specifications, directives, official standard regulations and/or expected results.

Reconstitute the vial with 9 ml of sterile diluent (50% Ethanol:water) in aseptic conditions and add it to 500 ml of Oxford Agar base cooled to 50°C.

Do not overheat once suplemented.

Pour the complete medium into Petri dishes and, once solidified on a flat surface, spread the plates by streaking or spyral method. Incubate the plates in aerobic atmosphere at  $37 \pm 1^{\circ}$ C for  $44 \pm 4h$ .

Incubation times longer than those mentioned above or different incubation temperatures may be requied depending on the sample or the specifications.

After incubation, enumerate all the colonies that have appeared onto the surface of the agar, observing any blackening of the medium due to esculin hydrolysis, typical for Listeria strains.

Presumptive isolation of Listeria must be confirmed by further microbiological and biochemical tests.

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## **Quality control**

## Physical/Chemical control

Color: Yellow- orangey

## **Microbiological control**

Reconstitute 1 vial as indicated in COMPOSITION; shake and dissolve completely

Add 1 vial to 500 ml of medium base. DO NOT HEAT once supplemented.

Inoculate 30-300 CFU (productivity) 1.000-10.000 CFU (selectivity)

Analytical methodology according to ISO 11133:2014/A1:2018; A2:2020.

Distribute the complete medium, cooled to 50 °C, into 90 mm plates

Incubate according instructions for complete medium indicated in COMPOSITION.

Aerobiosis. Incubation at 37 ± 1 °C, reading after 24/44 ± 4 h

### Microorganism

L. monocytogenes ATCC® 13932, WDCM 00021 Escherichia coli ATCC® 25922, WDCM 00013 Enterococcus faecalis ATCC® 29212, WDCM 00087 L. monocytogenes ATCC® 35152, WDCM 00109

#### Growth

Good - Esculin Positive reaction Inhibited Inhibited Good - Esculin Positive reaction

### Sterility control

Add 5mL of the sample to 100 mL of TSB and to 100 mL Thioglycollate. Incubation 48 h at 30-35 °C and 48 h at 20-25 °C: NO GROWTH. Check at 7 days after incubation in same conditions.

## **Bibliography**

- · ATLAS, R.M. (1993) Handbook of Microbiological Media. CRC Press. Boca Raton. Florida.
- · CURTIS, G.D, R.G. MITCHELL, A.F. KING & E.J. GRIFFIN (1989) A selective differential medium for the isolation of Listeria monocytogenes. Letters Appl. Microbiol. 8:95-98.
- . ISO 11133:2014/ Adm 1:2018. Microbiology of food, animal feed and water. Preparation, production, storage and performance testing of culture media.
- · ISO 11290 standard (1996) Microbiology of food and animal feeding stuff. Horizontal method for the detection and enumeration of Listeria monocytogenes. Part 1 - Detection method. Part 2 - Enumeration method.
- · ISO 11290-1:2017 Standard. Microbiology of the food chain. Horizontal method for the detection and enumeration of Listeria monocytogenes and for Listeria spp.- Part 1: Detection Method
- · ISO 11290-2:2017 Standard. Microbiology of the food chain. Horizontal method for the detection and enumeration of Listeria monocytogenes and for Listeria spp.- Part 2: Enumeration Method
- · VANDERZANT, C. & D.F. SPLITTSTOESSER (1992) Compendium of methods for the microbiological examination of foods. APHA. Washington DC.
- . UNE-EN ISO 11133 (2014). Microbiología de los alimentos para consumo humano, alimentación animal y agua.-Preparación, producción, conservación y ensayos de rendimiento de los medios de cultivo.

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